

review

Natural killer cells: In health and disease



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Abstract Natural killer (NK) cells constitute our bodies' frontline defense system, guarding against tumors and launching attacks against infections. The activities of NK cells are regulated by the interaction of various receptors expressed on their surfaces with cell surface ligands. While the role of NK cells in controlling tumor activity is relatively clear, the fact that they are also linked to various other disease conditions is now being highlighted. Here, we present an overview of the role of NK cells during normal body state as well as under diseased state. We discuss the possible utilization of these powerful cells as immunotherapeutic agents in combating diseases such as asthma, autoimmune diseases, and HIV-AIDS. This review also outlines current challenges in NK cell therapy.

KEYWORDS: Natural killer (NK) cells; NK cells and cancer; NK cells and HIV1; NK cells and autoimmunity; Immunotherapy

Pathogen invasion in our body is counteracted by both adaptive and innate immune cells. The adaptive immune system is represented by B and T cells. B cells play a major role in the humoral immune response, whereas T cells are primarily involved in cell-mediated immune responses. The innate immune system consists of cells and proteins that play a crucial part in the initiation and subsequent activation of the adaptive immune system. They also participate in the removal of pathogens that have been targeted by an adaptive immune response. The main components of the innate immune system are physical epithelial barriers, phagocytic leukocytes, dendritic cells, and natural killer (NK) cells.

NK cells are crucial components of the innate immune system and, as the name suggests, they do not require pre-stimulation to perform their effector functions.¹ Morphologically, they are characterized as large, granular, bone marrow-derived lymphocytes and phenotypically, they are defined as CD56⁺ CD3⁻ in humans. They represent 10% of the cells in the total peripheral blood mononuclear cell (MNC) population of circulating human lymphocytes and they comprise the third largest population of lymphocytes following B and T cells. They are also found

in the peritoneal cavity, spleen, liver, lung, lymph nodes, thymus, and in uterus during gestation.

NK-CELL DEVELOPMENT

It is generally accepted that NK cells develop primarily in the bone marrow, similar to B cells and myeloid origin cells. However, recent studies have shown that NK cells can also develop in lymph nodes and liver.^{2,3} The generation of NK cells from hematopoietic stem cells (HSC) is a continuous process. In the first step, the HSC shows commitment towards NK-cell lineage. NK-cell precursors (NKP) have been identified in the hematopoietic population, which differentiates into NK cells but not to other lineages. This process is followed by phenotypic and functional NK-cell maturation. In the final step, NK cells undergo homeostasis. Several transcription factors as well as soluble and membrane factors have been identified that regulate NK-cell development and maturation. Transcription factors involved in the generation of NKP include Ets-1, Id2, Ikaros and PU.1.^{4,5} Maturation of immature NK cells is regulated by Gata-3 and IRF-2 and functional differentiation of matured NK cells involves CEBP- γ , MEF and MITF. The

cytokine IL-15 has been shown to be essential for NK-cell development, homeostasis and survival.⁶ Studies by Freud and Ferlazzo have implicated the role of T cell derived IL-2 in the cytolytic functional maturation of NK cells.^{2,7}

NK CELL FUNCTION

Natural killer cells have diverse biological functions which include recognizing and killing virally-infected and neoplastic cells. Circulating NK cells are mostly in their resting phase but activation by cytokines leads to infiltration of these cells into most tissues that contain pathogen-infected or malignant cells.^{8,9} NK cells also have an immunoregulatory role as they secrete several cytokines, such as interferon (IFN)- γ , following their ligand interaction with cell-surface receptors. Human NK cells can be classified into two subsets, depending on their immunophenotype and function: CD56^{dim} and CD56^{bright}. CD56^{dim} constitutes 90% of the total NK cell population in peripheral blood and these express a low-affinity receptor for the constant region of immunoglobulin G, Fc γ RIIIa (CD16).¹⁰ Functionally, these have high cytotoxic activity. Approximately 10% of NK cells belong to the CD56^{bright} subset and they are mostly involved in the production of cytokines.

The NK cells in the secondary lymphoid tissue such as tonsils, lymph nodes, and spleen are different from the NK cells in the peripheral blood as these are activated by dendritic cells and they secrete cytokines such as interferon, which stimulate a more efficient killing response by the T cells.^{7,11}

NK-cell functioning is controlled by a wide range of receptors that are expressed on the cell surface. These receptors are either inhibitory or activating in nature. The family of inhibitory receptors consists of the killer immunoglobulin-like receptors (KIR) or Ig-like receptors (CD158), the C type lectin receptors (CD94-NKG2A) and leukocyte inhibitory receptors (LIR1, LAIR-1). Activating receptors are the natural cytotoxicity receptors (NKp46, NKp44), C type lectin receptors (NKG2D, CD94-NKG2C), and Ig-like receptors (2B4). A particular NK cell typically expresses two to four inhibitory receptors in addition to an array of activation receptors. As different NK cells express different combinations of inhibitory or activating receptors, there is sizeable heterogeneity within the NK-cell population. It is for this reason that NK cells are considered to have the ability to respond to a variety of stimuli and to participate in immune responses under different pathological conditions.

NK-cell cytotoxicity is tightly regulated by a balance between activating and inhibitory signals. The inhibitory NK-cell receptors recognize self-MHC class I molecule, and this prevents NK-cell activation. This explains self tolerance and prevention of host cell killing. It was earlier discovered that NK cells are activated when they encounter cells which lack self-MHC class I molecule. This is known as the 'missing-self' hypothesis.¹² Moreover, NK cells can discriminate between normal host cells and infected or abnormal cells by recognition of MHC class I molecules. Virally infected cells and tumor cells often downregulate MHC class I expression to escape recognition by cytotoxic T lymphocytes (CTL), but this results in their vulnerability towards NK-cell attack. In this condition, activation receptors are no longer suppressed and they induce potent stimulatory signals, therefore tipping the balance in favor of NK-cell activation.^{13,14} This condition is often referred to as induced-self recognition.

Once the target is recognized by NK cells, their cytotoxic ability is mainly mediated via two predominant pathways. A membrane-disrupting protein, perforin, and a family of structurally related serine proteases, granzymes, are secreted by exocytosis, which jointly induce apoptosis of the target cell. In the second pathway, a caspase-dependent apoptosis takes place involving the association of death receptors (e.g. Fas/CD95) on target cells with their equivalent ligands such as FasL, and tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) on NK cells, resulting in caspase-dependent apoptosis. Antibody dependent cellular cytotoxicity (ADCC) can also be a mechanism of killing of tumor cells by NK cells as they express a low-affinity Fc receptor for IgG, Fc γ RIII (CD16).

NK CELLS DURING VARIOUS PHASES OF HUMAN LIFE

Human NK cells are present in fetal liver as early as gestational week 6, and in fetal spleen at gestational week 15.¹⁵ Although fetal liver NK cells are known to have the ability to kill target cells, they are hyporesponsive compared to adult NK cells. This indicates that fetal liver NK cells are functionally immature. It is likely that during the first trimester, NK-cell development is under a dynamic phase. This is followed by a steady phase in the second trimester. The transition from fetal NK-cell development to a more adult-like NK-cell state occurs in the third trimester.¹⁶ Activity of NK cells in newborns is considerably lower compared to adults, as has been shown in

a study by Kaplan *et al.*¹⁷ This inactivation has been accounted for by the lack of activation of pre NK cells *in vivo*. This deficiency may be associated with the susceptibility of newborns to certain viral infections such as herpes viruses, against which NK cells are considered to be the first line of defense.

During pregnancy, peripheral blood NK cells are found to be suppressed both in terms of number and activity.^{18,19} NK cells, besides being present in the peripheral blood, can also be detected in the uterus. This set of NK cells is known as uterine natural killer (uNK) cells. uNK cells do not express CD16, unlike peripheral blood NK cells, but they express CD94 and secrete cytokines such as MIP1 α , GM-CSF, CSF1 and IFN- γ .²⁰ It has been suggested that uNK cells are either a distinct subpopulation of peripheral blood NK cells or they could have arisen by tissue-specific differentiation. At the implantation site, uNK cells are the most prominent leukocytes present. During the proliferative phase of the menstrual cycle, uNK cells are few in number. Their level rises significantly during the secretory phase and continues to remain high during early gestation. At 20 weeks' gestation NK cells decrease and are absent in term deciduas.^{21,22} They play an important role in controlling trophoblast invasion and express receptors that interact with ligands expressed on trophoblast.²³ NK cells are an important regulator of spiral artery remodeling and maintenance of decidual integrity.²⁴ Fu *et al.* demonstrated that NK cells regulate pathogenic T helper 17 (Th17) cells at the maternal-fetal interface and thus promote immune tolerance and maintenance of pregnancy.²⁵ They are also responsible for switching the pro-inflammatory state to the anti-inflammatory state in the endometrium by down-regulating the expression of a soluble decoy receptor (ST2L) receptor on their surface molecule, which binds to IL-33. Imbalance in IL33/ST2 activation can lead to recurrent pregnancy loss.²⁶

ROLE IN VARIOUS DISEASE CONDITIONS

The involvement of NK cells has been recognized in various disease conditions. As mentioned previously, one of the primary functions of NK cells is immuno-surveillance of our body. Several *in vitro* studies on mammalian cells, including human cells, and also *in vivo* studies in mice and rats prove that NK cells recognize tumor cells as targets. They control tumor growth and metastasis diffusion *in vivo*. Tumor immuno-surveillance role of NK cells has also been implicated in controlling the growth of B cell

lymphomas that spontaneously arise in mice lacking both perforin and β 2-microglobulin.²⁷ An epidemiologic survey of 11-year follow-up shows a link between low NK cell activity in peripheral blood and increased cancer risk in adults.²⁸ The role of NK cells as host immunity has also been studied in various cases of infections by flaviviruses, such as Japanese encephalitis virus, yellow fever virus, dengue virus, tick-borne encephalitis virus and West Nile virus (WNV). Their role in viral hepatitis, influenza virus and HIV-1 infection is also well documented in several studies.^{29–34} Similarly, their role in protecting against respiratory infection by bacteria, viruses such as respiratory syncytial viruses (RSV), and influenza has been elaborately described in murine studies.^{35–37} NK cells are assumed to be a major determinant of the development of viral-associated asthma.

In most cases, the role of NK cells is found to be either disease controlling or disease enhancing. For example, in asthma, NK cells contribute towards the progress of T cell mediated allergic airway response during allergen specific sensitization phase.^{38,39} Existing evidence also suggests that NK cells are involved in resolving acute allergic airway inflammation.⁴⁰ Peripheral blood of asthmatic patients shows enhanced NK-cell activity which decreases upon antigen challenge.⁴¹ This suggests that NK cells migrate from circulation towards lungs and lymphoid organs.^{41,42} Similarly, in human autoimmune diseases, changes have been observed in circulating blood NK cells in terms of quantitative as well as qualitative parameters. In many instances of autoimmune disease, a reduction in number of NK cells along with decreased cytotoxic function has been observed.^{43,44} *In vivo* studies using experimental autoimmune encephalomyelitis (EAE), which is an animal model of multiple sclerosis (MS), show increased severity and mortality when NK cells are depleted prior to disease induction. EAE animals have cellular infiltration, CNS inflammation, and demyelination.^{45–47} It is therefore hypothesized that NK cells are involved in the control of autoimmune disease conditions. Clinical trials on MS patients suggest low frequency and activity of NK cells in the peripheral blood but this cannot be ascertained as these studies were conducted using variable methods and low patient sample size.^{48–50} On the other hand, the cytotoxicity of NK cells can augment an autoimmune disease. Auto reactive NK cells can lead to the destruction of cells in a target organ. In Type 1 diabetes, NK cells have been found in pancreatic islets only during infection or inflammation, and not under healthy, non-diseased conditions.⁵¹ Preclinical data also

suggest that NK cells are involved in the development of Type 1 diabetes.^{52,53} Some studies on Type 1 diabetes patients show that NK cells are either decreased or their function is impaired.^{54–56} In rheumatoid arthritis (RA), tissue NK cells have disease promoting functions. In 2005, Laszlo reported that patients with RA have NK-cell accumulation in their synovial fluid.⁵⁷ The NK-cell subset, CD56^{bright}, found here, secretes more IFN γ compared with blood NK cells from the same patients.⁵⁸ However, in systemic lupus erythematosus (SLE), patients show a variable and moderate reduction of NK-cell numbers along with reduced CD4⁺CD25⁺ Treg cells.^{59,60} The function of NK cells is downregulated in these patients and there is a shift from the CD56^{dim} population to the CD56^{bright} subset.^{61,62} It is also indicated that NK cells in these patients have a reduced cytotoxic effect.⁶³ This deficiency of NK cells corresponds with clinical conditions such as nephritis and thrombopenia during SLE. The abnormality in NK-cell number and function could therefore play a role in the inflammatory condition. In other words, NK cells play a protective or disease controlling role to prevent SLE.

It is evident that NK cells act on cells during disease condition either through their receptors or due to the interactions of cytokines. They are known to attack tumor cells when the expression of MHC class I molecules is absent or downregulated. Upregulation of NKG2D ligands on tumor cells can also make them susceptible to NK-cell attack. Most cancer cells engage the NK cell's activating receptors, which triggers its natural kill response. Members of the NK-cell receptor family also contribute towards the defense mechanism against viruses. Infection of mouse or human cells with flaviviruses is known to increase cell-surface expression of MHC class I on infected cells, as evidenced in WNV infection, and therefore they evade NK-cell mediated killing.^{64,65} In HIV1 infection, although no specific NK-cell receptors have been identified that recognize HIV1 infected cells, there is a remarkable increase in inhibitory receptors and a decrease in number of activating receptors like NKp30, NKp46 on NK cells.^{66,67} *In vivo* condition has shown that NK cell ligand HLA-B Bw4-801 and its receptor KIR 3DS1 form an association resulting in the inhibition of HIV-1 replication and the killing of target cells by NK cells.^{32,34} This leads to a decrease in activity of NK cells during HIV1 infection. The abnormality of NK-cell functioning during HIV-1 infection can be credited to viral proteins. HIV-1 gp41, gp120, Nef and Tat have been proven to downregulate NK-cell activity by various

mechanisms.^{68–71} Evidence of NK-cell receptor involvement is also known in diabetes. Gur *et al.* recently demonstrated that NKp46, the activating NK-cell receptor, binds to an unknown ligand on pancreatic β cells effectively killing them, due to the degranulation of NK cells in mice as well as humans.^{51,54} The study concluded that NKp46 is essential for the development of Type 1 diabetes. In humans, this ligand is expressed constitutively in both the young and in adults. But the fact that not all humans become diabetic in spite of possessing the ligand that makes β cells subject to NK-cell attack is because NK cells are not commonly found in the healthy pancreas. In another study, patients with long-standing Type 1 diabetes showed a remarkable low expression of NKp30 and NKp46 activating receptors in their blood in comparison to those of the control group. Also, the expression of NKG2D was found to be reduced relative to the control and irrespective of disease duration. Long-standing patients also displayed reduced perforin mRNA expression.⁷² Consistent with these results, a decreased lysis activity by the NK cells was observed by Lorini *et al.* in patients with long-standing diabetes.⁷³ The reduction in NK-cell activity in these diabetic patients is thought to be a consequence rather than a cause.

An important role is played by cytokines and chemokines which act in conjunction with NK cells to tackle various diseased conditions. IL-12 and IL-18, NK activating cytokines active during late NK-cell differentiation, have been demonstrated to synergistically enhance cytotoxicity against tumor targets and IFN- γ production by NK cells. IFN- γ induces type I immune response and directly acts on cancer cells. IL-12, IL-18 and IFN- γ are also known to have a pro-atherogenic effect.⁷⁴ In response to certain viral infections, IFN- α/β is produced, enhancing the NK-cell mediated cytotoxicity and leading to the killing of the viral infected cells. Moreover, many key pathways related to antiviral functions are activated by IFN- γ . IL-21, another cytokine binding the common γ -chain (shared with IL-2, IL-4, IL-7, IL-9, and IL-15), has been demonstrated to favor the onset of the most cytotoxic CD56^{dim}CD16⁺ NK cell subset and to enhance its cytotoxicity. Tumor Necrosis Factor (TNF) is another factor produced by NK cells and which is known to mediate antiviral and immunoregulatory effects. Chemokines produced by NK cells such as MIP-1 α are capable of promoting inflammatory processes. In some cases, IL-10, also produced by NK cells, is known to be anti-inflammatory which inhibits Dendritic Cells (DCs). NK cells can lessen

the effect of antigen presentation by Antigen Presenting Cells (APCs) and reduce T cell proliferation.⁷⁵ These cells are normally observed to have accumulated at the site of immunization, and they generate cytokines which are involved in the pathogenesis of allergic inflammation. NK cells are indicated to generate IFN- γ , TNF- α , GM-CSF and MIP-1 α upon stimulation with IgE, and also demonstrate cytotoxicity against IgE coated target cells in a Fc γ RIII dependent manner.⁷⁶

NK cells interact with various other immune cells in our body both in normal conditions as well as during pathological conditions. In normal and asthmatic lungs, lung resident dendritic cells and macrophages are known to form synapses with NK cells leading to generation of NK derived cytokines and effector molecules involved in local immunity, and at the same time can regulate allergic disease severity.⁷⁷ The dynamic nature of cytokine and cellular profile of the microenvironment can influence the development of specific NK subtypes which may lead to conversion from a pro-inflammatory to a pro-resolution NK subtype.⁷⁸ If there is any disturbance or defect in this process, then it may lead to more severe inflammation and eventually to airway damage. DCs are known to cross-talk with NK cells through production of cytokines

such as IL-12 and IL-18 as well as through cell–cell interactions to promote NK-cell activity against tumors. Crosstalk between NK cells and DCs may be disrupted during HIV1 infection, although this mechanism is not clear. Moreover, it is possible for NK-cell mediated lysis of virally infected cells to be a source of apoptotic bodies for uptake of DCs, which may promote DC maturation and viral antigen presentation to T cells.⁷⁹ Another important immune system component, the macrophages, are known to increase the anti-tumor and anti-infection activity of NK cells through their crosstalk.^{80–82} Treg cells, part of the adaptive immunity, are also known to interact with NK cells and mostly control their activity during various disease conditions. Treg cells have been seen to suppress NK cells via IL-21 mediation in autoimmune disease conditions.⁸³ Similarly, in patients with gastrointestinal, colon and prostate cancers, a high level of Treg cells has been associated with a reduced number of NK cells along with reduced functionality.^{84–86} An *in vitro* study has shown that Treg cells from hepatocellular carcinoma patients inhibit NK-cell killing ability. However, during pregnancy, NK cells, along with Treg cells, contribute towards the creation of tolerant conditions for the fetus, and any change in that leads to complications (Table 1).

Table 1. NK cells in disease conditions.

Disease	Role of NK cells	Status of NK cell during disease condition	Possible therapeutic approach	References
Cancer	Immuno-surveillance	Low activity	Adoptive NK cell transfer and enhancement of activatory receptors	27,28,87–96
Viral infection	Immuno-protection	Low number and activity, shift from CD56 ^{dim} to CD56 ^{bright}	Adoptive NK cell transfer, genetically engineered HIV1 specific NK cells receptors, CCR5 deficient hESC-NK cell transfer	29–37
Asthma	Contribution to IgE mediated immune-response, resolution of airway inflammations	Migration from circulation to lung and lymphoid organs	Adoptive NK cell transfer, <i>in vitro</i> or <i>in vivo</i> expansion of specific NK cell subsets	38–42
Type 1 diabetes	Disease enhancing	Migration from blood to pancreas? Low expression of NKp30, NKp46 and NKG2D, low perforin in blood NK cells	Targeting NKp46 receptor to reduce auto-destruction of β -cells	51–54,72,73
Rheumatoid arthritis	Disease controlling or enhancing?	Low number and activity in blood, increase in synovial fluid	Blocking inhibitory NKG2A (to enhance) or RANKL and M-CSF (to control)	44,57,58,97
Systemic lupus erythematosus	Disease controlling	Low number and activity, CD56 ^{bright} increase, low perforin	Adoptive NK cell transfer	59–63

Abbreviations: hESC – human embryonic stem cells.

THERAPEUTIC APPLICATIONS OF NK CELLS IN VARIOUS DISEASE CONDITIONS

NK cells play a crucial role in attacking tumor cells in our bodies, and are considered a promising tool for cancer therapy. Treatment range over the past two decades has included IL-2 administration to activate the endogenous NK cells or to adoptively transfer IL-2 activated NK cells.⁸⁷⁻⁹¹ Autologous NK-cell therapy has been experimented on for the treatment of renal cell carcinoma, malignant glioma, and metastatic breast cancer. However, it was soon recognized that autologous adoptive NK-cell therapy may have certain drawbacks and thus may not be efficacious. The drawback is mostly attributed to the inhibition of NK cells by self-MHC I molecules expressed on the tumor cells. This has led to the use of allogeneic NK cell therapy in trials. In a pioneering study, Ruggeri *et al.* demonstrated that alloreactive NK cells given to patients with acute myelogenous leukemia (AML) could eliminate relapse, graft rejection, and protect them against graft-vs-host disease (GvHD).⁹² Later, adoptive cellular transfer of allogeneic NK cells from haploidentical donors was also attempted for treatment of renal cell carcinoma, metastatic melanoma, refractory Hodgkin's disease, and refractory AML.⁹³ They were also found to be useful against several solid tumors such as neuroblastoma, renal, colon, gastric, and ovarian cancers.^{94,95} The trials concluded that NK-cell transfer was safe and efficacious. Similar trials were also conducted recently in patients with recurrent metastatic breast and ovarian cancer.⁹⁶ The allogeneic NK cells have the advantage of being derived from healthy donors and have more cytotoxic activity. Moreover, NK cells do not induce GvHD, unlike T cells.

As discussed in the earlier section, the role of NK cells has been established not only in cancer but also in various other disease conditions. Adoptive NK cell therapy can thus be explored for diseases such as asthma, multiple sclerosis, diabetes, arthritis, etc. The effectiveness of NK cells in controlling HIV-1 infection has already been demonstrated in *in vitro* and *in vivo* experiments.^{31,33} NK cell therapy can be applied to patients who are refractory to standard highly active antiretroviral therapy (HAART). Besides the option of using NK cells for adoptive transfers, understanding the role of NK cells and their receptors can open up other strategies to treat diseases. For example, during the developmental stages of Type 1 diabetes, the activation of NK cells can be prevented by the administration of specific anti-

bodies for blocking the NKp46 activation receptor. Similarly, in rheumatoid arthritis where the role of NK cells can possibly be protective or disease-enhancing, therapy can be considered accordingly. Inhibitory receptor NKG2A can be blocked, which will stimulate NK cells and thus control the disease. Where NK cells enhance the disease condition, the blocking of RANKL (receptor activator of NF κ B ligand) and M-CSF (macrophage colony-stimulating factor), factors which mediate osteoclastogenesis and bone destruction, can help.⁹⁷

For the purpose of therapeutic applications, allogeneic NK cells can be sourced from umbilical cord blood (UCB), adult donor lymphapheresis products, or even from NK-cell lines such as NK-92. Recently, studies have shown successful *in vitro* derivation of functional NK cells from human embryonic stem cell (hESC) and induced pluripotent stem cell (iPSC).⁹⁸⁻¹⁰⁰ hESC and iPSC-derived NK cells have demonstrated potent anti-tumorigenic and anti HIV activity, and are phenotypically similar to those of peripheral blood origin. Moreover, they are considered superior to UCB-derived NK cells because they have higher levels of KIR expression, thus making them more potent. Pluripotent cell-derived NK cells can therefore be an unlimited source for the adoptive transfer of NK cells to treat a range of diseases. However, safety of hESC and iPSC-derived NK cells in terms of potential tumorigenicity needs to be determined before they can be utilized in the clinical set up.

The application of NK cells as immunotherapeutic agent requires several technical developments. NK cells need to be isolated and expanded in sufficient numbers for them to act as effector cells. Moreover, the activity of NK cells needs to be enhanced for better efficacy. Expansion of NK cells has been attempted using cytokines such as IL-2 and IL-15.^{101,102} These two cytokines can also help increase the survivability of the NK cells.¹⁰³ IL-2 is also thought to potentiate the cytotoxic ability of NK cells. Co-culturing NK cells with accessory cells such as irradiated Epstein Barr Virus (EBV) transformed lymphoblastoid cells, HFWT (a Wilm's tumor derived cell line), and K562 has been reported to enhance NK cell proliferation.¹⁰⁴⁻¹⁰⁶ Activation of NK cells can be achieved by various genetic engineering techniques to augment activating signals and also to downregulate inhibitory signals.¹⁰⁷⁻¹¹¹ Similarly, the specificity of NK cells can be increased through genetic modification approaches such as the use of chimeric antigen receptors (CARs)¹¹²⁻¹¹⁴ (Table 1).

CONCLUSION

Research to date has helped us gain an understanding of NK cell biology in terms of function and role of their receptor interactions. Their task in attacking tumor cells is now well established. However, a clearer picture to determine their specific roles in diseases such as asthma, diabetes, and rheumatoid arthritis is still desired. Further investigation is required to understand the interactions of NK cells with other cells of the immune system such as T cells, dendritic cells, and macrophages. But there is no doubt that NK cells will emerge as major players in the area of

cancer treatments, viral infections, including HIV/AIDS, autoimmune diseases, and asthma in the coming decade. The immediate future may see the use of NK cell therapy in combination with chemotherapy, radiotherapy, and surgery for cancer. More focus should be placed on establishing techniques for the isolation and expansion of these cells in their required numbers.

CONFLICT OF INTEREST

We have no conflict of interest to declare.

REFERENCES

- Smyth MJ, Hayakawa Y, Takeda K, Yagita H. New aspects of natural-killer-cell surveillance and therapy of cancer. *Nat Rev Cancer* 2002;2(11):850–61.
- Freud AG, Becknell B, Roychowdhury S, Mao HC, Ferketich AK, Nuovo GJ, et al. A human CD34(+) subset resides in lymph nodes and differentiates into CD56^{bright} natural killer cells. *Immunity* 2005;22(3):295–304.
- Andrews DM, Smyth MJ. A potential role for RAG-1 in NK cell development revealed by analysis of NK cells during ontogeny. *Immunol Cell Biol* 2010;88(2):107–16.
- Barton K, Muthusamy N, Fischer C, Ting CN, Walunas TL, Lanier LL, et al. The Ets-1 transcription factor is required for the development of natural killer cells in mice. *Immunity* 1998;9(4):555–63.
- Boggs SS, Trevisan M, Patrene K, Georgopoulos K. Lack of natural killer cell precursors in fetal liver of Ikaros knockout mutant mice. *Nat Immun* 1998;16(4):137–45.
- Ma A, Koka R, Burkett P. Diverse functions of IL-2, IL-15, and IL-7 in lymphoid homeostasis. *Annu Rev Immunol* 2006;24:657–79.
- Ferlazzo G, Pack M, Thomas D, Paludan C, Schmid D, Strowig T, et al. Distinct roles of IL-12 and IL-15 in human natural killer cell activation by dendritic cells from secondary lymphoid organs. *Proc Natl Acad Sci U S A* 2004;101(47):16606–11.
- Fogler WE, Volker K, McCormick KL, Watanabe M, Ortaldo JR, Willtrout RH. NK cell infiltration into lung, liver, and subcutaneous B16 melanoma is mediated by VCAM-1/AVLA-4 interaction. *J Immunol* 1996;156(12):4707–14.
- Glas R, Franksson L, Une C, Eloranta ML, Ohlén C, Örn A, et al. Recruitment and activation of natural killer (NK) cells *in vivo* determined by the target cell phenotype. An adaptive component of NK cell-mediated responses. *J Exp Med* 2000;191(1):129–38.
- Caligiuri MA. Human natural killer cells. *Blood* 2008;112(3):461–9.
- Vitale M, Della Chiesa M, Carlomagno S, Romagnani C, Thiel A, Moretta L, et al. The small subset of CD56^{bright}CD16⁺ natural killer cells is selectively responsible for both cell proliferation and interferon-gamma production upon interaction with dendritic cells. *Eur J Immunol* 2004;34(6):1715–22.
- Kärre K, Ljunggren HG, Piontek G, Kiessling R. Selective rejection of H-2-deficient lymphoma variants suggests alternative immune defence strategies. *Nature* 1986;319(6055):675–8.
- Malnati MS, Lusso P, Ciccone E, Moretta A, Moretta L, Long EO. Recognition of virus-infected cells by natural killer cell clones is controlled by polymorphic target cell elements. *J Exp Med* 1993;178(3):961–9.
- Bauer S, Groh V, Wu J, Steinle A, Phillips JH, Lanier LL, et al. Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. *Science* 1999;285(5428):727–9.
- Phillips JH, Hori T, Nagler A, Bhat N, Spits H, Lanier LL. Ontogeny of human natural killer (NK) cells: fetal NK cells mediate cytolytic function and express cytoplasmic CD3 epsilon, delta proteins. *J Exp Med* 1992;175(4):1055–66.
- Ivarsson MA, Loh L, Marquardt N, Kekäläinen E, Berglin L, Björkström NK. Differentiation and functional regulation of human fetal NK cells. *J Clin Invest* 2013;123(9):3889–901.
- Kaplan J, Shope TC, Bollinger RO, Smith J. Human newborns are deficient in natural killer activity. *J Clin Immunol* 1982;2(4):350–5.
- Sacks G, Sargent I, Redman C. An innate view of human pregnancy. *Immunol Today* 1999;20(3):114–8.
- Yovel G, Shakhar K, Ben-Eliyahu S. The effects of sex, menstrual cycle, and oral contraceptives on the number and activity of natural killer cells. *Gynecol Oncol* 2001;81(2):254–62.
- Moffett-King A. Natural killer cells and pregnancy. *Nat Rev Immunol* 2002;2(9):656–63.
- Bulmer JN, Morrison L, Longfellow M, Ritson A, Pace D. Granulated lymphocytes in human endometrium: histochemical and immunohistochemical studies. *Hum Reprod* 1991;6(6):791–8.
- Trundle A, Moffett A. Human uterine leukocytes and pregnancy. *Tissue Antigens* 2004;63(1):1–12.
- Moffett-King A, Entrican G, Ellis S, Hutchinson J, Bainbridge D. Natural killer cells and reproduction. *Trends Immunol* 2002;23(7):332–3.
- Tirado-González I, Barrientos G, Freitag N, Otto T, Thijssen VL, Moschansky P, et al. Uterine NK cells are critical in shaping DC immunogenic functions compatible with pregnancy progression. *PLoS One* 2012;7(10):e46755.
- Fu B, Li X, Sun R, Tong X, Ling B, Tian Z, et al. Natural killer cells promote immune tolerance by regulating inflammatory TH17 cells at the human maternal-fetal interface. *Proc Natl Acad Sci U S A* 2013;110(3):E231–40.
- Salker MS, Nautiyal J, Steel JH, Webster Z, Sućurović S, Nicou M, et al. Disordered IL-33/ST2 activation in decidualizing stromal cells prolongs uterine receptivity in women with recurrent pregnancy loss. *PLoS One* 2012;7(12):e52252.
- Street SE, Hayakawa Y, Zhan Y, Lew AM, MacGregor D, Jamieson AM, et al. Innate immune surveillance of spontaneous B cell lymphomas by natural killer cells and gamma/delta T cells. *J Exp Med* 2004;199(6):879–84.
- Imai K, Matsuyama S, Miyake S, Suga K, Nakachi K. Natural cytotoxic activity of peripheral-blood lymphocytes and cancer incidence: an 11-year follow-up study of a general population. *Lancet* 2000;356(9244):1795–9.
- Lunemann S, Malone DF, Hengst J, Port K, Grabowski J, Deterding K, et al. Compromised function of natural killer cells in acute and chronic viral hepatitis. *J Infect Dis* 2014;209(9):1362–73.
- Leung KN, Ada GL. Induction of natural killer cells during murine influenza virus infection. *Immunobiology* 1981;160(3–4):352–66.
- Guo H, Kumar P, Moran TM, Garcia-Sastre A, Zhou Y, Malarkannan S. The functional impairment of natural killer cells during influenza virus infection. *Immunol Cell Biol* 2009;87(8):579–89.
- Alter G, Martin MP, Teigen N, Carr WH, Suscovich TJ, Schneidewind A, et al. Differential natural killer cell-mediated inhibition of HIV-1 replication based on distinct KIR/HLA subtypes. *J Exp Med* 2007;204(12):3027–36.
- Iannello A, Debbeche O, Samarani S, Ahmad A. Antiviral NK cell responses in HIV infection: I. NK cell receptor genes as determinants of HIV resistance and progression to AIDS. *J Leukoc Biol* 2008;84(1):1–26.
- Martin MP, Gao X, Lee JH, Nelson GW, Detels R, Goedert JJ, et al. Epistatic interaction between KIR3DS1 and HLA-B delays the progression to AIDS. *Nat Genet* 2002;31(4):429–34.
- Stein-Streilein J, Bennett M, Mann D, Kumar V. Natural killer cells in mouse lung: surface phenotype, target preference, and response to local influenza virus infection. *J Immunol* 1983;131(6):2699–704.
- Stein-Streilein J, Guffee J. *In vivo* treatment of mice and hamsters with antibodies to asialo GM1 increases morbidity and mortality to pulmonary influenza infection. *J Immunol* 1986;136(4):1435–41.
- Nogusa S, Ritz BW, Kassim SH, Jennings SR, Gardner EM. Characterization of age-related changes in natural killer cells during primary influenza infection in mice. *Mech Ageing Dev* 2008;129(4):223–30.
- Wingett D, Nielson CP. Divergence in NK cell and cyclic AMP regulation of T cell CD40L expression in asthmatic subjects. *J Leukoc Biol* 2003;74(4):531–41.
- Ple C, Barrier M, Amniai L, Marquillies P, Bertout J, Tscipoulos A, et al. Natural killer cells accumulate in lung-draining lymph nodes and regulate airway eosinophilia in a murine model of asthma. *Scand J Immunol* 2010;72(2):118–27.

40. Haworth O, Cernadas M, Levy BD. NK cells are effectors for resolvin E1 in the timely resolution of allergic airway inflammation. *J Immunol* 2011;186(11):6129–35.

41. Jira M, Antosova E, Vondra V, Strejcek J, Mazakova H, Prazakova J. Natural killer and interleukin-2 induced cytotoxicity in asthmatics. I. Effect of acute antigen-specific challenge. *Allergy* 1988;43(4):294–8.

42. Culley FJ. Natural killer cells in infection and inflammation of the lung. *Immunology* 2009;128(2):151–63.

43. Yabuhara A, Yang FC, Nakazawa T, Iwasaki Y, Mori T, Koike K, et al. A killing defect of natural killer cells as an underlying immunologic abnormality in childhood systemic lupus erythematosus. *J Rheumatol* 1996;23(1):171–7.

44. Aramaki T, Ida H, Izumi Y, Fujikawa K, Huang M, Arima K, et al. A significantly impaired natural killer cell activity due to a low activity on a per-cell basis in rheumatoid arthritis. *Mod Rheumatol* 2009;19(3):245–52.

45. Zhang B, Yamamura T, Kondo T, Fujiwara M, Tabira T. Regulation of experimental autoimmune encephalomyelitis by natural killer (NK) cells. *J Exp Med* 1997;186(10):1677–87.

46. Xu W, Fazekas G, Hara H, Tabira T. Mechanism of natural killer (NK) cell regulatory role in experimental autoimmune encephalomyelitis. *J Neuroimmunol* 2005;163(1–2):24–30.

47. Hao J, Liu R, Piao W, Zhou Q, Vollmer TL, Campagnolo DI, et al. Central nervous system (CNS)-resident natural killer cells suppress Th17 responses and CNS autoimmune pathology. *J Exp Med* 2010;207(9):1907–21.

48. Benczur M, Petrányi GG, Pálffy G, Varga M, Tálas M, Kotsy B, et al. Dysfunction of natural killer cells in multiple sclerosis: a possible pathogenetic factor. *Clin Exp Immunol* 1980;39(3):657–62.

49. Oger J, Kastrukoff LF, Li DK, Paty DW. Multiple sclerosis: in relapsing patients, immune functions vary with disease activity as assessed by MRI. *Neurology* 1988;38(11):1739–44.

50. Braakman E, van Tunen A, Meager A, Lucas CJ. Natural cytotoxic activity in multiple sclerosis patients: defects in IL-2/interferon gamma-regulatory circuit. *Clin Exp Immunol* 1986;66(2):285–94.

51. Gur C, Porgador A, Elboim M, Gazit R, Mizrahi S, Stern-Ginossar N, et al. The activating receptor Nkp46 is essential for the development of type 1 diabetes. *Nat Immunol* 2010;11(2):121–8.

52. Maruyama T, Watanabe K, Takei I, Kasuga A, Shimada A, Yanagawa T, et al. Anti-asialo GM1 antibody suppression of cyclophosphamide-induced diabetes in NOD mice. *Diabetes Res* 1991;17(1):37–41.

53. Maruyama T, Watanabe K, Yanagawa T, Kasatani T, Kasuga A, Shimada A, et al. The suppressive effect of anti-asialo GM1 antibody on low-dose streptozotocin-induced diabetes in CD-1 mice. *Diabetes Res* 1991;16(4):171–5.

54. Gur C, Enk J, Kassem SA, Suissa Y, Magenheim J, Stolovich-Rain N, et al. Recognition and killing of human and murine pancreatic beta cells by the NK receptor Nkp46. *J Immunol* 2011;187(6):3096–103.

55. Brauner H, Elemans M, Lemos S, Broberger C, Holmberg D, Flodström-Tullberg M, et al. Distinct phenotype and function of NK cells in the pancreas of nonobese diabetic mice. *J Immunol* 2010;184(5):2272–80.

56. Poulton LD, Smyth MJ, Hawke CG, Silveira P, Shepherd D, Naidenko OV, et al. Cytometric and functional analyses of NK and NKT cell deficiencies in NOD mice. *Int Immunol* 2001;13(7):887–96.

57. Pazmany L. Do NK cells regulate human autoimmunity? *Cytokine* 2005;32(2):76–80.

58. Dalbeth N, Callan MF. A subset of natural killer cells is greatly expanded within inflamed joints. *Arthritis Rheum* 2002;46(7):1763–72.

59. Erkeller-Yuksel FM, Lydyard PM, Isenberg DA. Lack of NK cells in lupus patients with renal involvement. *Lupus* 1997;6(9):708–12.

60. Erkeller-Yüsel F, Hulstaart F, Hannel I, Isenberg D, Lydyard P. Lymphocyte subsets in a large cohort of patients with systemic lupus erythematosus. *Lupus* 1993;2(4):227–31.

61. Hervier B, Beziat V, Haroche J, Mathian A, Lebon P, Ghillani-Dalbin P, et al. Phenotype and function of natural killer cells in systemic lupus erythematosus: excess interferon- γ production in patients with active disease. *Arthritis Rheum* 2011;63(6):1698–706.

62. Schepis D, Gunnarsson I, Eloranta ML, Lampa J, Jacobson SH, Kärre K, et al. Increased proportion of CD56^{bright} natural killer cells in active and inactive systemic lupus erythematosus. *Immunology* 2009;126(1):140–6.

63. Park YW, Kee SJ, Cho YN, Lee EH, Lee HY, Kim EM, et al. Impaired differentiation and cytotoxicity of natural killer cells in systemic lupus erythematosus. *Arthritis Rheum* 2009;60(6):1753–63.

64. Lobigs M, Blanden RV, Müllbacher A. Flavivirus-induced up-regulation of MHC class I antigens; implications for the induction of CD8⁺ T-cell-mediated autoimmunity. *Immunol Rev* 1996;152:5–19.

65. Ye J, Zhu B, Fu ZF, Chen H, Cao S. Immune evasion strategies of flaviviruses. *Vaccine* 2013;31(3):461–71.

66. Mavilio D, Benjamin J, Daucher M, Lombardo G, Kottliil S, Planta MA, et al. Natural killer cells in HIV-1 infection: dichotomous effects of viremia on inhibitory and activating receptors and their functional correlates. *Proc Natl Acad Sci U S A* 2003;100(25):15011–6.

67. De Maria A, Fogli M, Costa P, Murdaca G, Puppo F, Mavilio D, et al. The impaired NK cell cytolytic function in viremic HIV-1 infection is associated with a reduced surface expression of natural cytotoxicity receptors (Nkp46, Nkp30 and Nkp44). *Eur J Immunol* 2003;33(9):2410–8.

68. Huber M, Trkola A. Humoral immunity to HIV-1: neutralization and beyond. *J Intern Med* 2007;262(1):5–25.

69. Cohen GB, Gandhi RT, Davis DM, Mandelboim O, Chen BK, Strominger JL, et al. The selective downregulation of class I major histocompatibility complex proteins by HIV-1 protects HIV-infected cells from NK cells. *Immunity* 1999;10(6):661–71.

70. Cerboni C, Neri F, Casartelli N, Zingoni A, Cosman D, Rossi P, et al. Human immunodeficiency virus 1 Nef protein downmodulates the ligands of the activating receptor NKG2D and inhibits natural killer cell-mediated cytotoxicity. *J Gen Virol* 2007;88(Pt 1):242–50.

71. Carroll IR, Wang J, Howcroft TK, Singer DS. HIV Tat represses transcription of the beta 2-microglobulin promoter. *Mol Immunol* 1998;35(18):1171–8.

72. Rodacki M, Svoren B, Butty V, Besse W, Laffel L, Benoist C, et al. Altered natural killer cells in type 1 diabetic patients. *Diabetes* 2007;56(1):17–85.

73. Lorini R, Moretta A, Valtorta A, d’Annunzio G, Cortona L, Vitali L, et al. Cytotoxic activity in children with insulin-dependent diabetes mellitus. *Diabetes Res Clin Pract* 1994;23(1):37–42.

74. Kleemann R, Zadelaar S, Kooistra T. Cytokines and atherosclerosis: a comprehensive review of studies in mice. *Cardiovasc Res* 2008;79(3):360–76.

75. Deniz G, Erten G, Küçüksezer UC, Kocacik D, Karagiannidis C, Aktas E, et al. Regulatory NK cells suppress antigen-specific T cell responses. *J Immunol* 2008;180(2):850–7.

76. Arase N, Arase H, Hirano S, Yokosuka T, Sakurai D, Saito T. IgE-mediated activation of NK cells through Fc gamma RIII. *J Immunol* 2003;170(6):3054–8.

77. Wehner R, Dietze K, Bachmann M, Schmitz M. The bidirectional crosstalk between human dendritic cells and natural killer cells. *J Innate Immun* 2011;3(3):258–63.

78. Walzer T, Vivier E. G-protein-coupled receptors in control of natural killer cell migration. *Trends Immunol* 2011;32(10):486–92.

79. Altfeld M, Fadda L, Frelata D, Bhardwaj N. DCs and NK cells: critical effectors in the immune response to HIV-1. *Nat Rev Immunol* 2011;11(3):176–86.

80. Nedvetzki S, Sowinski S, Eagle RA, Harris J, Vély F, Pende D, et al. Reciprocal regulation of human natural killer cells and macrophages associated with distinct immune synapses. *Blood* 2007;109(9):3776–85.

81. Lapaque N, Walzer T, Méresse S, Vivier E, Trowsdale J. Interactions between human NK cells and macrophages in response to Salmonella infection. *J Immunol* 2009;182(7):4339–48.

82. Zhou Z, Zhang C, Zhang J, Tian Z. Macrophages help NK cells to attack tumor cells by stimulatory NKG2D ligand but protect themselves from NK killing by inhibitory ligand Qa-1. *PLoS One* 2012;7(5):e36928.

83. Liu R, Van Kaer L, La Cava A, Price M, Campagnolo DI, Collins M, et al. Autoreactive T cells mediate NK cell degeneration in autoimmune disease. *J Immunol* 2006;176(9):5247–54.

84. Ghiringhelli F, Ménard C, Terme M, Flament C, Taieb J, Chaput N, et al. CD4⁺CD25⁺ regulatory T cells inhibit natural killer cell functions in a transforming growth factor-beta-dependent manner. *J Exp Med* 2005;202(8):1075–85.

85. Doubrovina ES, Doubrovin MM, Vider E, Sisson RB, O’Reilly RJ, Dupont B, et al. Evasion from NK cell immunity by MHC class I chain-related molecules expressing colon adenocarcinoma. *J Immunol* 2003;171(12):6891–9.

86. Wu JD, Higgins LM, Steinle A, Cosman D, Haugk K, Plymate SR. Prevalent expression of the immunostimulatory MHC class I chain-related molecule is counteracted by shedding in prostate cancer. *J Clin Invest* 2004;114(4):560–8.

87. Rosenberg SA. Interleukin-2 and the development of immunotherapy for the treatment of patients with cancer. *Cancer J Sci Am* 2000;6(Suppl. 1):S2–7.

88. Rosenberg SA. Immunotherapy of cancer by systemic administration of lymphoid cells plus interleukin-2. *J Biol Response Mod* 1984;3(5):501–11.

89. Rosenberg SA, Lotze MT, Muul LM, Chang AE, Avis FP, Leitman S, et al. A progress report on the treatment of 157 patients with advanced cancer using lymphokine-activated killer cells and interleukin-2 or high-dose interleukin-2 alone. *N Engl J Med* 1987;316(15):889–97.

90. Hayes RL, Koslow M, Hiesiger EM, Hymes KB, Hochster HS, Moore EJ, et al. Improved long term survival after intracavitary interleukin-2 and lymphokine-activated killer cells for adults with recurrent malignant glioma. *Cancer* 1995;76(5):840–52.

91. Keilholz U, Scheibenbogen C, Brado M, Georgi P, Maclachlan D, Brado B, et al. Regional adoptive immunotherapy with interleukin-2 and lymphokine-activated killer (LAK) cells for liver metastases. *Eur J Cancer* 1994;30A(1):103–5.

92. Ruggeri L, Capanni M, Urbani E, Perruccio K, Shlomchik WD, Tosti A, et al. Effectiveness of donor natural killer cell alloreactivity in mismatched hematopoietic transplants. *Science* 2002;295(5562):2097–100.

93. Miller JS, Soignier Y, Panoskaltis-Mortari A, McNearney SA, Yun GH, Fautsch SK, et al. Successful adoptive transfer and *in vivo* expansion of human haploidentical NK cells in patients with cancer. *Blood* 2005;105(8):3051–7.

94. Castriconi R, Dondero A, Corrias MV, Lanino E, Pende D, Moretta L, et al. Natural killer cell-mediated killing of freshly isolated neuroblastoma

- cells: critical role of DNAX accessory molecule-1-poliovirus receptor interaction. *Cancer Res* 2004;64(24):9180–4.
- 95.** Carlsten M, Björkström NK, Norell H, Bryceson Y, van Hall T, Baumann BC, et al. DNAX accessory molecule-1 mediated recognition of freshly isolated ovarian carcinoma by resting natural killer cells. *Cancer Res* 2007;67(3):1317–25.
- 96.** Geller MA, Cooley S, Judson PL, Ghebre R, Carson LF, Argenta PA, et al. A phase II study of allogeneic natural killer cell therapy to treat patients with recurrent ovarian and breast cancer. *Cytotherapy* 2011;13(1):98–107.
- 97.** Teitelbaum SL. Bone resorption by osteoclasts. *Science* 2000;289(5484):1504–8.
- 98.** Woll PS, Martin CH, Miller JS, Kaufman DS. Human embryonic stem cell-derived NK cells acquire functional receptors and cytolytic activity. *J Immunol* 2005;175(8):5095–103.
- 99.** Woll PS, Grzywacz B, Tian X, Marcus RK, Knorr DA, Vermeris MR, et al. Human embryonic stem cells differentiate into a homogeneous population of natural killer cells with potent *in vivo* antitumor activity. *Blood* 2009;113(24):6094–101.
- 100.** Ni Z, Knorr DA, Clouser CL, Hexum MK, Southern P, Mansky LM, et al. Human pluripotent stem cells produce natural killer cells that mediate anti-HIV-1 activity by utilizing diverse cellular mechanisms. *J Virol* 2011;85(1):43–50.
- 101.** Trinchieri G, Matsumoto-Kobayashi M, Clark SC, Sehra J, London L, Perussia B. Response of resting human peripheral blood natural killer cells to interleukin 2. *J Exp Med* 1984;160(4):1147–69.
- 102.** Carson WE, Fehniger TA, Haldar S, Eckhert K, Lindemann MJ, Lai CF, et al. A potential role for interleukin-15 in the regulation of human natural killer cell survival. *J Clin Invest* 1997;99(5):937–43.
- 103.** Becknell B, Caligiuri MA. Interleukin-2, interleukin-15, and their roles in human natural killer cells. *Adv Immunol* 2005;86:209–39.
- 104.** Berg M, Lundqvist A, McCoy Jr P, Samsel L, Fan Y, Tawab A, et al. Clinical-grade ex vivo-expanded human natural killer cells up-regulate activating receptors and death receptor ligands and have enhanced cytolytic activity against tumor cells. *Cytotherapy* 2009;11(3):341–55.
- 105.** Harada H, Watanabe S, Saijo K, Ishiwata I, Ohno T. A Wilms tumor cell line, HFWT, can greatly stimulate proliferation of CD56⁺ human natural killer cells and their novel precursors in blood mononuclear cells. *Exp Hematol* 2004;32(7):614–21.
- 106.** Robertson MJ, Cameron C, Lazo S, Cochran KJ, Voss SD, Ritz J. Costimulation of human natural killer cell proliferation: role of accessory cytokines and cell contact-dependent signals. *Nat Immun* 1996–1997;15(5):213–26.
- 107.** Boyiadzis M, Memon S, Carson J, Allen K, Szczepanski MJ, Vance BA, et al. Up-regulation of NK cell activating receptors following allogeneic hematopoietic stem cell transplantation under a lymphodepleting reduced intensity regimen is associated with elevated IL-15 levels. *Biol Blood Marrow Transplant* 2008;14(3):290–300.
- 108.** Altvater B, Landmeier S, Pscherer S, Temme J, Schweer K, Kailayangiri S, et al. 2B4 (CD244) signaling by recombinant antigen-specific chimeric receptors costimulates natural killer cell activation to leukemia and neuroblastoma cells. *Clin Cancer Res* 2009;15(15):4857–66.
- 109.** Imai C, Iwamoto S, Campana D. Genetic modification of primary natural killer cells overcomes inhibitory signals and induces specific killing of leukemic cells. *Blood* 2005;106(1):376–83.
- 110.** Godal R, Bachanova V, Gleason M, McCullar V, Yun GH, Cooley S, et al. Natural killer cell killing of acute myelogenous leukemia and acute lymphoblastic leukemia blasts by killer cell immunoglobulin-like receptor-negative natural killer cells after NKG2A and LIR-1 blockade. *Biol Blood Marrow Transplant* 2010;16(5):612–21.
- 111.** Fujisaki H, Kakuda H, Shimasaki N, Imai C, Ma J, Lockett T, et al. Expansion of highly cytotoxic human natural killer cells for cancer cell therapy. *Cancer Res* 2009;69(9):4010–7.
- 112.** Schirmann T, Pecher G. Specific targeting of CD33(+) leukemia cells by a natural killer cell line modified with a chimeric receptor. *Leuk Res* 2005;29(3):301–6.
- 113.** Müller T, Uherek C, Maki G, Chow KU, Schimpf A, Klingemann HG, et al. Expression of a CD20-specific chimeric antigen receptor enhances cytotoxic activity of NK cells and overcomes NK-resistance of lymphoma and leukemia cells. *Cancer Immunol Immunother* 2008;57(3):411–23.
- 114.** Kruschinski A, Moosmann A, Poschke I, Norell H, Chmielewski M, Seliger B, et al. Engineering antigen-specific primary human NK cells against HER-2 positive carcinomas. *Proc Natl Acad Sci U S A* 2008;105(45):17481–6.